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Collective migration plays a pivotal role in numerous epithelial tissues, under both physiological and pathological conditions. Recent evidence suggests a correlation between the activation waves of extracellular signal-regulated kinase (ERK) and cell deformation, which together orchestrate collective cell migration. Additionally, emerging evidence indicates that localized changes in matrix curvature can influence the migration of individual epithelial cells (known as curvotaxis). However, the precise mechanism by which local changes in curvature modulate ERK propagation and contribute to coordinating collective movement remains unclear. Here we utilize a photopolymerization technique to create well-defined corrugation patterns of varying wavelengths in soft hydrogels, thereby mimicking the multiscale curvature found in human tissues. To elucidate the role of ERK waves in directing cell migration, we employ Förster resonance energy transfer (FRET)-based biosensor in MDCK cells, conducting time-lapse experiments lasting 24 hours on culture substrates featuring both flat and corrugated regions. Our findings reveal distinct flow directions within the same epithelial tissue, transitioning from flat to corrugated hydrogels. Furthermore, we demonstrate that ERK activity is significantly heightened in corrugated regions, suggesting a mechanism that facilitates collective cell migration on curved substrates.

ERK activity couples to polarization to direct collective cell migration

ERK activity is quantified by FRET

- adherens junctions







hydrogel



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